

INFORMATION TO USERS

The most advanced technology has been used to photograph and reproduce this manuscript from the microfilm master. UMI films the text directly from the original or copy submitted. Thus, some thesis and dissertation copies are in typewriter face, while others may be from any type of computer printer.

The quality of this reproduction is dependent upon the quality of the copy submitted. Broken or indistinct print, colored or poor quality illustrations and photographs, print bleedthrough, substandard margins, and improper alignment can adversely affect reproduction.

In the unlikely event that the author did not send UMI a complete manuscript and there are missing pages, these will be noted. Also, if unauthorized copyright material had to be removed, a note will indicate the deletion.

Oversize materials (e.g., maps, drawings, charts) are reproduced by sectioning the original, beginning at the upper left-hand corner and continuing from left to right in equal sections with small overlaps. Each original is also photographed in one exposure and is included in reduced form at the back of the book.

Photographs included in the original manuscript have been reproduced xerographically in this copy. Higher quality 6" x 9" black and white photographic prints are available for any photographs or illustrations appearing in this copy for an additional charge. Contact UMI directly to order.

U·M·I

University Microfilms International
A Bell & Howell Information Company
300 North Zeeb Road, Ann Arbor, MI 48106-1346 USA
313/761-4700 800/521-0600

PREVIEW

Order Number 9080115

**Evaluation of progress from selection for cold and freeze
tolerance in maize**

Eichelberger, Kevin D., Ph.D.

The University of Nebraska - Lincoln, 1990

U·M·I
300 N. Zeeb Rd.
Ann Arbor, MI 48106

PREVIEW

EVALUATION OF PROGRESS FROM SELECTION FOR
COLD AND FREEZE TOLERANCE IN MAIZE

by

Kevin D. Eichelberger

A DISSERTATION

Presented to the Faculty of
The Graduate College in the University of Nebraska
In Partial Fulfillment of Requirements
For the Degree of Doctor of Philosophy

Major: Agronomy

Under the Supervision of Professor Charles O. Gardner

Lincoln, Nebraska

May, 1990

DISSERTATION TITLE

Evaluation of Progress from Selection for Cold and Freeze

Tolerance in Maize

BY

Kevin D. Eichelberger

SUPERVISORY COMMITTEE:

APPROVED

DATE

C. O. Gardner
Signature

4/6/90

Dr. C. O. Gardner
Typed Name

Blain Johnson
Signature

6 April 90

Dr. B. E. Johnson
Typed Name

C. Y. Sullivan
Signature

4/6/90

Dr. C. Y. Sullivan
Typed Name

Paul Staswick
Signature

4/6/90

Dr. P. Staswick
Typed Name

Dermot P. Coyne
Signature

4/6/90

Dr. D. P. Coyne
Typed Name

Signature

Typed Name



EVALUATION OF PROGRESS FROM SELECTION FOR
COLD AND FREEZE TOLERANCE IN MAIZE

Kevin D. Eichelberger, Ph.D.

University of Nebraska, 1990

Advisor: C. O. Gardner

In 1972, a full-sib recurrent selection program for improved cold tolerance was initiated in the maize (Zea mays L.) synthetic population NS, a Nebraska version of Iowa Stiff-Stalk Synthetic. Selection was based primarily on percent emergence and visual vigor-rating under controlled (11.1/4.4°C day/night) conditions. Visual selection was also practiced for agronomic traits. After an early freeze in the 1976 nursery, selection was initiated to form a freeze-tolerant subpopulation.

The first objective of this study was to evaluate progress from selection for cold tolerance, and to examine changes in agronomic traits. Materials evaluated included cycles 0, 4, 8, and 12 of the cold-tolerant population (NS-CT-C0, C4, C8, and C12) and cycles 8 and 12 of the freeze-tolerant subpopulation (NS-CT-FT-C8 and C12). In general, much more improvement in cold tolerance was noted in the growth chamber than in the field. Grain yield of NS-CT-C12 and NS-CT-FT-C12 was increased by 1.10 and 0.85 Mg ha⁻¹, respectively, relative to NS-CT-C0. The NS-CT and NS-CT-FT selections also flowered earlier, were lower in plant and

ear height, and had lower grain moisture at harvest than NS-CT-C0.

The second objective was to measure genotypic variation for cold tolerance in the aforementioned entries, under cool conditions in the growth chamber and in an early planting in the field. In general, NS-CT-C0 displayed more genotypic variation for cold tolerance than did the selected populations. Selection appeared to have acted in eliminating highly susceptible genotypes in the early cycles. Genetic associations among cold tolerance traits were generally low, indicating that they are controlled by different genetic systems. Correspondence between performance in the growth chamber and in the field was low.

The third objective was to evaluate freeze tolerance of the above entries in the field and in the growth chamber. No appreciable level of freeze tolerance was noted in either environment. Nor was any differential regrowth after freezing noted. Much more damage was caused to plants by temperatures of -3.0 versus -2.5°C .

ACKNOWLEDGEMENTS

The author wishes to express his appreciation to his major advisor, Dr. C. O. Gardner, for his guidance, advice, and time during the research for and preparation of this manuscript.

To Dr. B. E. Johnson for serving as co-chairperson of the supervisory committee, and to Drs. C. Y. Sullivan, D. P. Coyne, and P. Staswick for serving on the committee, the author would also like to express his sincerest gratitude.

Appreciation is expressed to Drs. W. A. Compton and M. A. Thomas-Compton for helpful discussions during the course of this study.

Appreciation is also expressed to technicians, secretaries and fellow graduate students for help, encouragement, and advice, and to the Department of Agronomy for financial support.

Finally, to his wife, Debbie, and his son, David, the author would like to express his gratitude for their patience and understanding during the research for and preparation of this manuscript.

TABLE OF CONTENTS

| | PAGE |
|--|------|
| INTRODUCTION. | 1 |
| LITERATURE REVIEW | 6 |
| Physiological Aspects of Chilling Sensitivity | |
| in Maize. | 6 |
| Primary Physiological Events During Chilling | |
| Stress. | 7 |
| Photosynthesis | 11 |
| Respiration. | 15 |
| Water Relations. | 16 |
| Other Physiological Considerations | 18 |
| Cold Tolerance and Stage of Development. | 21 |
| Seed Maturation | 21 |
| Germination | 23 |
| Heterotrophic Growth. | 25 |
| Autotrophic Growth. | 29 |
| Flowering to Physiological Maturity | 30 |
| Genetics of Maize Cold Tolerance | 33 |
| Selection for Cold Tolerance in Maize. | 38 |
| Physiological and Genetic Aspects of Freeze | |
| Tolerance in Maize. | 41 |
| MATERIALS AND METHODS | 46 |
| Selection Program. | 46 |
| Evaluation of Cold Tolerance and Agronomic | |
| Traits of Bulk Populations. | 50 |

TABLE OF CONTENTS (Continued)

| | PAGE |
|---|------|
| MATERIALS AND METHODS (Continued) | |
| Genetic Study of Cold Tolerance in Original and Selected Populations. | 53 |
| Evaluation of Freeze Tolerance of Bulk Populations | 56 |
| RESULTS AND DISCUSSION. | 60 |
| Field and Growth Chamber Evaluation of Bulks of Original and Selected Populations. | 60 |
| Field and Growth Chamber Evaluation of Full-sib Families from Original and Selected Populations | 80 |
| Evaluation of Freeze Tolerance of Original and Selected Populations. | 111 |
| CONCLUSIONS | 119 |
| LITERATURE CITED. | 122 |
| APPENDIX. | 134 |

Improving tolerance to adverse environmental conditions is an important goal of many maize (Zea mays L.) breeders. Cold tolerance, defined as the ability to germinate, emerge, and grow relatively normally in cool, wet soils early in the growing season, is a trait that is desirable under a number of circumstances. Early planting of maize by producers in temperate regions is deemed advantageous in order to distribute the workload more evenly over the growing season. Early planting also makes earlier flowering possible, thereby reducing risk of heat and drought stress during pollination, and may allow for more efficient use of early season precipitation. Higher grain yields are often obtained with early planting (Pendleton and Egli, 1969; Benoit et al., 1965; Imholte and Carter, 1987; Eckert, 1984). Early planting may also result in lower grain moisture at harvest (Imholte and Carter, 1987; Eckert, 1984), and increased stalk strength (Pendleton and Egli, 1969; Imholte and Carter, 1987). In high latitude or altitude regions, early planting is often essential for maximum production and for avoidance of damaging frosts before maturity. However, early planting of maize often results in poor stand establishment and reduced seedling vigor, which may decrease the yield potential of the crop. Cold tolerance would be especially useful in conjunction with reduced tillage or no-till cultural practices, where plant residues left on the soil

surface result in lower soil temperatures (Mock and Erbach, 1977) and higher moisture levels (Blevins et al., 1971).

A prerequisite for genetic improvement of any trait is the existence of heritable genetic variation. Numerous researchers have detected significant genetic variation for cold tolerance in maize. Grogan (1970) concluded that cold tolerance was conditioned primarily by additive effects, but also found dominance, epistasis, and maternal effects to be important. Pesev (1970) concluded that maternal effects were important and that the genetic control of cold tolerance is complex. Mock and Eberhart (1972) observed significant genetic variation for percent emergence and rate of emergence among S_1 lines from two breeding populations adapted to the U.S. Corn Belt. Eagles and Hardacre (1979a) found dominance genetic effects to be more important than additive effects for percent emergence and time to emergence at 10°C in a highland tropical maize population. In a generation means analysis involving three "cool season" and three "warm season" inbred lines, McConnell and Gardner (1979a) found additive, dominance, and epistatic gene effects to be important for germination at a constant 7.2°C , as well as for percent emergence and seedling vigor rating in the field. These and other studies indicate that sufficient genetic variation exists to improve maize cold tolerance via selection.

Recurrent selection for cold tolerance in maize has been attempted. Mock and Bakri (1976) evaluated several

cycles of selection for cold tolerance in two U.S. Corn Belt-adapted populations. Selection was based on an index integrating percent emergence, rate of emergence, and seedling dry weight. Two cycles of recurrent selection increased percent emergence and seedling dry weight by 8.4% and 0.6 dg plant⁻¹, respectively, in one population (BSSS13(SCT)). The other population (BSS2(SCT)) showed no response after three cycles of selection. Rate of emergence showed no response to selection in either population. Hoard and Crosbie (1985) evaluated five cycles of selection in BSSS13(SCT) and BSSS2(SCT) in the field. Percent emergence increased at a rate of 2.4 and 2.2% cycle⁻¹ in the two respective populations. Increases in seedling dry weight and vigor rating, but not in rate of emergence, were reported for both populations. McConnell and Gardner (1979b) practiced recurrent selection for rapid germination at a constant 7.2°C in two U.S. Corn Belt-adapted populations. Four cycles of selection resulted in significant increases in percent germination of 8.8 and 9.9% cycle⁻¹ for the two populations in the laboratory at a constant 7.2°C. Little improvement was noted in percent emergence or vigor in early planted field experiments. This lack of response in the field was attributed to mild spring weather during the 2-year test period.

Plants in the field may also occasionally be subjected to frost in late spring. Frost can result in considerable damage to leaves or even death of the seedling. Even when

plants are not killed, frost damage may seriously affect the yield potential of the crop. Arny and Upper (1973) found that frost-damaged plants in the field yielded 29 percent less than their undamaged counterparts. Frost-damaged plants also had delayed maturity and higher grain moisture compared to undamaged plants.

The study of genetic aspects of freeze tolerance in maize has not received much attention. Dhillon et al. (1988) observed significant genotypic differences among 64 maize inbred lines for freezing injury in the growth chamber and the field. Gardner et al. (1987) reported that a population which had undergone several cycles of recurrent selection for cold and freeze tolerance was considerably more tolerant of freeze damage than the original population. However, Brar et al. (1987) found no significant genotypic variation in maize for tolerance to freezing temperatures. Miedema (1982) concluded that genetic variation for freeze tolerance in maize is low.

In 1972, a recurrent selection program for improved cold tolerance was initiated in a Nebraska version of Iowa Stiff-Stalk Synthetic at the University of Nebraska-Lincoln. Selection was based primarily on percent emergence and a visual rating of vigor after 30 days in the growth chamber under 11.1/4.4°C day/night conditions. After an early-season frost in the nursery in 1976, selection was initiated to form a freeze-tolerant subpopulation. The objectives of this study were, 1) to assess progress from

5
selection for cold tolerance, under conditions in the field and in the growth chamber; 2) to measure genetic variation and genetic relationships among cold tolerance traits in selected cycles of the cold- and freeze-tolerant populations in the field and in the growth chamber, and to compare results in the two environments; and 3) to evaluate freeze tolerance of original and selected populations in the field and in the growth chamber.

PREVIEW

Physiological Aspects of Chilling Sensitivity in Maize

Maize belongs to the chilling sensitive (CS) category of plant species, which includes those species which are sensitive to low, nonfreezing temperatures up to about 10-12°C (Lyons, 1973). Exposure of CS plants to chilling temperatures results in a number of visual injury symptoms, including reduced germination and emergence, retarded growth and development, wilting, chlorosis, and eventually, reduced vigor or even premature death (McWilliam, 1983; Blum, 1988). At the cellular and subcellular levels, exposure to chilling temperatures causes such disturbances as reduction or cessation of protoplasmic streaming, leakage of cell solutes and loss of turgor, disruption of cellular membranes, and inhibition of chloroplast development (Lyons, 1973). The extent and severity of damage depends on the intensity and duration of the stress, as well as on the developmental and physiological state of the plant. However, these visual symptoms are secondary manifestations of injury at a more basic level of organization.

The primary events or lesions induced by exposure of CS plants to chilling stress are not fully understood. Possible mechanisms of chilling injury include changes in the physical state of biological membranes; metabolic imbalances due to differences in the activation energies (E_a) between different enzymes or enzymatic systems; and

low-temperature-induced denaturation or dissociation of proteins, leading to loss of essential activity.

Primary Physiological Events During Chilling Stress

A widely accepted hypothesis explaining the damaging effect of chilling temperatures on CS plants holds that the primary event during chilling is a low-temperature-induced phase transition of the biological membranes. According to this hypothesis, membrane lipids of CS species undergo a thermotropic change from a fluid, liquid-crystalline state to a more rigid gel state at some critical low temperature. This hypothesis was first proposed by Lyons and Raison (1970) and was later presented in a more refined form by Lyons (1973). The hypothesis was based in part on the observation that Arrhenius plots of succinate oxidation in mitochondria isolated from three CS species showed a marked discontinuity (increase in slope) at the temperature that coincided with the onset of visual chilling injury symptoms in these plants. Succinate oxidase activity in mitochondria from three chilling resistant (CR) species exhibited no such discontinuity between 0 and 25°C (Lyons and Raison, 1970). Further evidence for this hypothesis was found in discontinuities of Arrhenius plots of other membrane-bound enzymatic systems at the same critical temperature (Lyons, 1973). It was postulated that a phase transition of the membranes would lead to an increase in the permeability of membranes and to an increase in activation energy (E_a) of many membrane-bound enzymes or

enzymatic systems. The increase in permeability would lead to an inability to maintain the proper ionic balance within the cell. The increased E_a of membrane-bound enzymes would lead to metabolic imbalances which in turn could eventually lead to a buildup of toxic metabolic intermediates. For example, it was hypothesized that the activity of mitochondrial membrane-bound enzymes of the TCA cycle would be more affected than would the soluble enzymes of glycolysis. This could lead to the accumulation of toxic levels of acetaldehyde and ethanol in plant cells (Lyons, 1973). If the chilling stress was alleviated in time, the plants would recover; if not, they would suffer irreversible injury or death.

The phase transition hypothesis has been tested in a large number of species using a variety of techniques, with mixed results. Earlier studies using electron spin resonance (ESR) spectroscopy seemed to indicate a reasonably close relationship between visual chilling sensitivity and occurrence of a phase transition of the membranes. However, the use of ESR spectroscopy to detect phase transitions has been criticized (Schreier *et al.*, 1978), as has the interpretation of the data obtained using this technique (Wolfe and Bagnall, 1979). More recent results have not been as consistent. For example, Raison and Orr (1986) used three different techniques - ESR, differential scanning calorimetry (DSC), and fluorescence techniques using the fluorescent probe trans-parinaric

acid- to examine chilling-induced phase changes in thylakoid polar lipids of oleander, mung bean, and tomato, all CS species. All three techniques detected a phase change at virtually the same temperature within each species. On the other hand, O'Neill and Leopold (1982) were unable to detect a phase transition above 0°C in phospholipid vesicles of mitochondrial membranes of CS soybeans using DSC. Similarly, Dalziel and Breidenbach (1982) were unable to detect a phase transition above 0°C with either DSC or ESR in two ecotypes of wild tomato which differed in chilling sensitivity.

The question of whether certain membrane systems of CS plants undergo a phase transition in the range of 0-12°C has not been fully resolved. Although it is an attractive hypothesis, its confirmation awaits further study.

An important part of the phase transition hypothesis is the expected increase in E_a of membrane-bound enzymes or enzymatic systems. Lyons and Raison (1970) hypothesized that a phase transition might affect lipid-protein interactions, thereby increasing the E_a of membrane-associated proteins. Their hypothesis was based on the observation that Arrhenius plots of the rate of succinate oxidation in mitochondria from CS tomato, cucumber fruit and sweet potato roots exhibited a deviation from linearity (interpreted as an increase in E_a) at about 9-12°C. No such deviation was found in mitochondria from CR cauliflower buds, potato tubers, or beet roots. This was

interpreted as a phase transition-induced partial inactivation of the enzyme. While an increase in E_a of a membrane-bound enzyme may not be harmful in itself, it was thought that such effects may result in metabolic imbalances, leading to a buildup of toxic metabolic intermediates (Lyons, 1973).

Other membrane-bound enzymes or enzymatic systems have also shown abrupt changes in E_a upon chilling. Murata et al. (1975) noted a break in the Arrhenius plot of photosynthetic activity above 0° C in chloroplasts from the CS blue-green alga Anacystis nidulans, which correlated with a phase transition event as detected by ESR. They found no temperature-induced break in Arrhenius plots of photosynthetic activity for CR spinach or lettuce. Shneyour et al. (1973) noted a temperature-induced break in the Arrhenius plot of photoreduction of NADP^+ in isolated chloroplasts from CS tomato and bean at about 12°C. No discontinuity in the Arrhenius plot was seen for CR lettuce or pea. More recently, Yoshida et al. (1986) noted a break at 6.7°C in the Arrhenius plot of Mg^{2+} -stimulated ATPase activity in highly purified plasma membrane fractions from CS mung bean. However, membrane-bound enzymes do not always exhibit sharp drops in E_a at temperatures where changes in the molecular ordering of lipids is indicated by ESR or other techniques. For example, Patterson et al. (1979) were unable to detect a break in the Arrhenius plot of O_2 uptake above 0°C in pollen membranes from CS tomato.

Furthermore, some CR species exhibit discontinuities in Arrhenius plots of membrane-bound enzymes at temperatures above 0°C, contrary to expectation. For example, Pomeroy and Andrews (1975) detected a break in the Arrhenius plot of plasma membrane ATPase activity at 11°C in wheat, a species which is completely chilling resistant.

In the past, an increase in E_a of membrane-bound enzymes has usually been ascribed to changes in the molecular ordering of membrane lipids. This change supposedly affects hydrophobic interactions between acyl portions of membrane lipids and hydrophobic amino acids of membrane proteins, resulting in a change in protein conformation and therefore a change (usually a decrease) in activity. However, chilling may have a direct effect on enzyme activity, independent of lipid-protein interactions. Conformational changes in proteins due to low temperatures (cold lability) are a distinct possibility (Lyons *et al.*, 1979). In fact, some enzymes which are not associated with membranes show breaks in Arrhenius plots of activity versus temperature. For example, Graham *et al.* (1979) noted a discontinuity in the Arrhenius plot for PEP carboxylase from CS tomato at about 10°C. PEP carboxylase from CR wheat showed no break in the Arrhenius plot above 0°C. Similarly, Uedan and Sugiyama (1976) noted an increased E_a of CS maize PEP carboxylase at 11-12°C.

Photosynthesis

Photosynthesis (Ps) in CS plants such as maize is

greatly inhibited at low temperatures. Duncan and Hesketh (1968) examined the effect of temperature on net photosynthetic rates of 21 races of maize and a modern single-cross hybrid (OH45 x K4). Average net photosynthetic rates were reduced 52 and 25%, respectively, at 15/10 and 18/13°C day/night temperatures compared to values measured at 30/25°C. Alberda (1969) observed negligible rates of Ps eight days after seedlings growing at 20°C were shifted to 10°C. Bird et al. (1977) also found photosynthetic rates in maize to be negligible at low temperature (13/10°C, day/night).

Several explanations are possible for the observed decrease in Ps at low temperatures. One is that certain photosynthetic enzymes are inhibited at some critical low temperature. As mentioned previously, Uedan and Sugiyama (1976) noted an increase in E_a of maize PEP carboxylase at 11-12°C. Stamp (1981a), on the other hand, found only small differences in PEP carboxylase activity of 28 maize inbreds when grown at low versus high (15/12.5°C versus 25/22.5°C day/night) temperature, while the activity of ribulosebisphosphate carboxylase declined 73% at the low temperature regime. Taylor et al. (1974) found that exposure of maize seedlings to 10°C for 3 days under high light reduced PEP carboxylase activity only slightly relative to 25°C-grown controls, while NADP-malate dehydrogenase and pyruvate dikinase activities declined by 39 and 50%, respectively. No loss in activity was observed

under lower light levels, however, indicating a light dependence of loss of activity.

Stomatal resistance to CO₂ uptake may also be important in reducing Ps at low temperatures. Low temperatures may reduce the hydraulic conductivity of root membranes of CS plants. In addition to reduced water uptake, the conductivity of the tonoplast of stomate guard cells may be reduced, rendering them less responsive to reduced leaf water potential at low temperatures. Bean (*Phaseolus vulgaris* L.) plants exposed to 5°C night temperatures had severely reduced levels of Ps the following day (Crookston *et al.*, 1974). This effect was paralleled by a decrease in leaf water potential and an increase in stomatal resistance to CO₂ uptake. When shoots (but not roots) were cooled, Ps, leaf water potential, and stomatal CO₂ resistance were similar to control levels.

Chilling CS plants in strong light enhances inhibition of Ps, and can lead to irreparable damage of the photosynthetic apparatus (Oquist and Martin, 1986). Long *et al.* (1983) studied the combined effect of high light intensity and low temperature on CO₂ assimilation rate in maize. Chilling young maize plants from 20°C to 5°C resulted in a 93% reduction in CO₂ assimilation rate. Upon return to 20°C, plants chilled in the dark regained 90% of their CO₂-assimilating capacity, compared to 50% for plants chilled in bright light.

Chlorosis is one of the most commonly reported