

INFORMATION TO USERS

While the most advanced technology has been used to photograph and reproduce this manuscript, the quality of the reproduction is heavily dependent upon the quality of the material submitted. For example:

- Manuscript pages may have indistinct print. In such cases, the best available copy has been filmed.
- Manuscripts may not always be complete. In such cases, a note will indicate that it is not possible to obtain missing pages.
- Copyrighted material may have been removed from the manuscript. In such cases, a note will indicate the deletion.

Oversize materials (e.g., maps, drawings, and charts) are photographed by sectioning the original, beginning at the upper left-hand corner and continuing from left to right in equal sections with small overlaps. Each oversize page is also filmed as one exposure and is available, for an additional charge, as a standard 35mm slide or as a 17"x 23" black and white photographic print.

Most photographs reproduce acceptably on positive microfilm or microfiche but lack the clarity on xerographic copies made from the microfilm. For an additional charge, 35mm slides of 6"x 9" black and white photographic prints are available for any photographs or illustrations that cannot be reproduced satisfactorily by xerography.

PREVIEW

8715844

Ayres, Nicola Marie

**INHERITANCE OF ORGANELLES IN SOMATIC CELL HYBRIDS AND
CYBRIDS OF NICOTIANA**

The University of Nebraska - Lincoln

Ph.D. 1987

**University
Microfilms
International**

300 N. Zeeb Road, Ann Arbor, MI 48106

PREVIEW

PREVIEW

PLEASE NOTE:

In all cases this material has been filmed in the best possible way from the available copy. Problems encountered with this document have been identified here with a check mark ✓.

1. Glossy photographs or pages ✓
2. Colored illustrations, paper or print _____
3. Photographs with dark background ✓
4. Illustrations are poor copy _____
5. Pages with black marks, not original copy ✓
6. Print shows through as there is text on both sides of page _____
7. Indistinct, broken or small print on several pages _____
8. Print exceeds margin requirements _____
9. Tightly bound copy with print lost in spine _____
10. Computer printout pages with indistinct print _____
11. Page(s) _____ lacking when material received, and not available from school or author.
12. Page(s) _____ seem to be missing in numbering only as text follows.
13. Two pages numbered _____. Text follows.
14. Curling and wrinkled pages _____
15. Dissertation contains pages with print at a slant, filmed as received ✓
16. Other _____

University
Microfilms
International

PREVIEW

INHERITANCE OF ORGANELLES
IN SOMATIC CELL HYBRIDS AND CYBRIDS
OF NICOTIANA

by

Nicola M. Ayres

A DISSERTATION

Presented to the Faculty of
The Graduate College in the University of Nebraska
In Partial Fulfillment of Requirements
For the Degree of Doctor of Philosophy

Major: Biological Sciences
(Genetics, Molecular, and Cellular Biology)

Under the Supervision of Professor David W. Galbraith

Lincoln, Nebraska

March, 1987

TITLE

Inheritance of Organelles in Somatic Cell Hybrids

and Cybrids of Nicotiana

BY

Nicola M. Ayres

APPROVED

DATE

David W. Galbraith

March 23, 1987

George E. Veomett

March 23, 1987

Sheldon Schuster

March 23, 1987

John Osterman

March 23, 1987

James Van Etten

March 23, 1987

SUPERVISORY COMMITTEE

GRADUATE COLLEGE

UNIVERSITY OF NEBRASKA

INHERITANCE OF ORGANELLES
IN SOMATIC CELL HYBRIDS AND CYBRIDS
OF NICOTIANA

Nicola M. Ayres, Ph.D.

University of Nebraska, 1987

Advisor: David W. Galbraith

In somatic cell hybrids and cybrids of Nicotiana, research has indicated the presence of a hierarchy of organelle dominance between various species. This putative hierarchy was based on results using biochemical selection of hybrids. I tested the possibility that biochemical selection pressure biased the results. Protoplasts of various alloplasmic substitutions of Nicotiana were fused together; heterologous fusion events were isolated using fluorescence-activated cell sorting. Leaves of plants regenerated from these fusion events were analyzed to identify the types of chloroplast and mitochondrial DNA present.

The results reported here indicate that the hierarchy of organelle dominance appears to be a real phenomenon, even when selective pressures are removed. Chloroplasts from Nicotiana undulata strongly outcompeted chloroplasts from N. tabacum. Chloroplasts from N. suaveolens and N. glauca equally competed with chloroplasts from N. tabacum. Analysis of mitochondrial DNA revealed that mitochondria segregated independently from

chloroplasts in the N. undulata - N. tabacum and N. glauca - N. tabacum fusions, but not in the N. suaveolens - N. tabacum fusions. After heterologous fusion events, mitochondrial DNA recombination was detected in some plants, but chloroplast DNA recombination was not observed.

In addition, male fertility was restored in a plant regenerated from a fusion between protoplasts of two cytoplasmic male sterile, alloplasmic substitutions. These alloplasmic substitutions contained the cytoplasm of N. suaveolens or N. plumbaginifolia, each in a N. tabacum nuclear background. Thus both nuclei were identical but each of the two protoplast types had the cytoplasm of either N. suaveolens or N. plumbaginifolia. The fertile plant contained N. plumbaginifolia chloroplast and mitochondrial DNA restriction patterns, and displayed a floral morphology intermediate between the N. plumbaginifolia alloplasmically substituted parent and fertile N. tabacum. The flowers of the fertile plant were only able to set seed when manually pollinated.

ACKNOWLEDGEMENTS

I would like to thank my advisor, Dr. David Galbraith, for critical discussion and helpful suggestions, as well as teaching me independence and scientific logic.

I would like to thank the members of my supervisory committee: Drs. James Van Etten, George Veomett, John Osterman, and Sheldon Schuster. Special thanks are due to Dr. John Osterman, whose mind contains more information than any book. It should be packaged and sold. Thanks for providing techniques, and advice long after being driven crazy by my questions.

Much gratitude is due to Kristi Harkins for magic hands on the flow cytometer, and many valuable suggestions.

I would like to thank C.S. Levings for the generous gift of mitochondrial DNA probes, J.F. Chaplin and G. Pitarelli for seed, and R.A. Jensen for the N. sylvestris cell line.

Special thanks to Glen Drohman for excellent care of the plants, even when I gave him a tobacco jungle, and to Chad Locke, for able and willing assistance.

Thanks to the Mad Argentinian, Claudio Afonso, for five years of friendship, advice, and being convenient to bully.

Special thanks to Carolyn Brown, Dr. Patricia Herman, Ann Krejci, Dr. Kit Wah Lee, Mildred Lonsdale, and all the others who gave me encouragement when the going got rough.

Finally I would like to thank my family for support in times of mental insanity (temporary, I hope).

TABLE OF CONTENTS

	Page
INTRODUCTION	1
LITERATURE REVIEW	2
Organelle Inheritance	2
Somatic Cell Fusion	6
Nuclear Segregation	7
Chloroplast Segregation	9
Mitochondrial Segregation.	10
Hierarchy of Organelle Dominance	11
Fluorescence-Activated Cell Sorting.	15
Experimental Plan - Organelle Inheritance	20
Cytoplasmic Male Sterility	21
Experimental Plan - Cytoplasmic Male Sterility	23
MATERIALS AND METHODS	24
I. Plant Material	24
II. Protoplast Fusion	26
A. Protoplast Isolation	26
B. Fluorescent Labeling	27
C. Protoplast Fusion	27
III. Fluorescent-Activated Cell Sorting	28
IV. Plant Regeneration	28

V.	DNA Content.	29
VI.	Pollen Germination	29
VII.	Chloroplast DNA Analysis	29
	A. Chloroplast Isolation.	29
	B. Chloroplast DNA Isolation	30
	C. Restriction Analysis	31
VIII.	Mitochondrial DNA Analysis	32
	A. Total DNA Isolation	32
	B. Restriction Analysis	32
	C. Southern Transfer	33
	D. Probes.	33
	E. Nick Translation.	34
	F. Hybridization	35
RESULTS	36
I.	Organelle Inheritance	36
	A. Fluorescence-Activated Cell Sorting	36
	B. Plant Regeneration	36
	C. DNA Content.	38
	D. Chloroplast DNA Analysis	41
	E. Mitochondrial DNA Analysis.	53
II.	Cytoplasmic Male Sterility	60
	A. Floral Analysis	60
	B. DNA Content.	62
	C. Chloroplast DNA Analysis	63
	D. Mitochondrial DNA Analysis.	63

DISCUSSION.	68
Organelle Inheritance	68
Cytoplasmic Male Sterility	76
CONCLUSIONS	79
LITERATURE CITED	80

PREVIEW

LIST OF FIGURES

	Page
Figure 1 Schematic Diagram of Cell Sorting . . .	18
Figure 2 Floral Morphology of Parental Plants . . .	25
Figure 3 Leaf Morphology	40
Figure 4 Chloroplast DNA Restriction Patterns of Regenerated Plants from tab/und/und- tab/tab/tab Fusions.	43
Figure 5 Chloroplast DNA Restriction Patterns of Regenerated Plants from tab/sua/sua- tab/tab/tab Fusions.	44
Figure 6 Chloroplast DNA Restriction Patterns of Regenerated Plants from tab/gla/gla- tab/tab/tab Fusions.	45
Figure 7 Autoradiograph and Sketch of Total DNA of Parental Plants Probed with ATPase Subunit Alpha	54
Figure 8 Autoradiograph and Sketch of Total DNA of Parental Plants Probed with ATPase Subunit Six	55
Figure 9 Autoradiograph of Total DNA of Regenerated Plants from tab/und/und-tab/tab/tab Fusions Probed with ATPase Subunit Alpha	56
Figure 10 Autoradiograph of Total DNA of Regenerated Plants from tab/und/und-tab/tab/tab Fusions Probed with ATPase Subunit Six	57
Figure 11 Autoradiograph of Total DNA of Regenerated Plants from tab/sua/sua-tab/tab/tab Fusions Probed with ATPase Subunit Alpha	58
Figure 12 Autoradiograph of Total DNA of Regenerated Plants from tab/gla/gla-tab/tab/tab Fusions Probed with ATPase Subunit Alpha	59

Figure 13	Flower Morphology of Parental and Fertile Plants	61
Figure 14	Chloroplast DNA Restriction Patterns of Regenerated Plants from tab/sua/sua-tab/pbg/pbg Fusions.	65
Figure 15	Autoradiograph of Total DNA of Regenerated Plants from tab/sua/sua-tab/pbg/pbg Fusions Probed with ATPase Subunit Alpha	67

PREVIEW

LIST OF TABLES

	Page
Table 1. Sorting Purity	37
Table 2. Tab(R)-Und(F) Analysis	39
Table 3. Tab(F)-Und(R) Analysis	47
Table 4. Tab(R)-Sua(F) Analysis	49
Table 5. Tab(F)-Sua(R) Analysis	50
Table 6. Tab(R)-Gla(F) Analysis	51
Table 7. Tab(F)-Gla(R) Analysis	52
Table 8. Seed Germination	63
Table 9. Pbg(R)-Sua(F) Analysis	66

ABBREVIATIONS

CMS, cytoplasmic male sterility; tab, *N. tabacum*; pbg, *N. plumbaginifolia*; sua, *N. suaveolens*; und, *N. undulata*; gla, *N. glauca*. The plants are referred to by the nomenclature: nuclear genome/chloroplast genome/mitochondrial genome. Hence tab/und/und refers to the alloplasmic substitution of und in a tab nuclear background. It possesses the chloroplasts and mitochondria of und and the tab. FACS, Fluorescence-activated cell sorting.

INTRODUCTION

When two different populations of isolated protoplasts are fused, the organelle populations are initially mixed. In regenerated plants, however, only one chloroplast and one mitochondrial type are usually detected. Some interaction between the organelles or between the organelles and the nuclei may govern this segregation. A putative hierarchy of organelle dominance has been implied from the results of protoplast fusion experiments, using Nicotiana protoplasts. In this hierarchy the chloroplasts of N. undulata strongly outcompete, of N. suaveolens slightly outcompete, and of N. glauca equally compete those of N. tabacum. The focus of the studies reported here was to determine whether the hierarchy of organelle dominance actually exists, or was a result of the experimental selective conditions that were employed. A second facet of this work was to study the restoration of male fertility in regenerated plants derived from a fusion of protoplasts of two cytoplasmically male sterile plants.

LITERATURE REVIEW

Organelle Inheritance

Early events in plant development include the interaction of organelles contributed by each of the parents. When fertilization occurs, the initial zygote is comprised of up to six different genomes: two nuclear, two mitochondrial, and two chloroplast genomes. In sexually produced hybrids, the number of genomes contributed to the zygote is determined by the type of organelle inheritance, either uniparental or biparental. In about sixty higher plants that have been classified for organelle inheritance, approximately two-thirds exhibit maternal inheritance; the remaining third has biparental plastid inheritance (Tilney-Bassett, 1984). Both types of plastid inheritance occur throughout the plant kingdom, indicating that the type of plastid inheritance has little evolutionary significance. The frequency with the change in type of plastid inheritance occurs implies that relatively few genes are responsible for plastid inheritance (Tilney-Bassett, 1984).

There are four types of plastid inheritance, based on when exclusion of plastids occurs (van Went, 1984). The first type is represented by Lycopersicon; the plastids are absent from the generative cells of the pollen grain. The vegetative cells contain plastids, but only the generative cell contributes to the

formation of the zygote. In some plants the plastids are present in the generative cells, but are eliminated during subsequent cell development leading to the formation of sperm cells. This type is characteristic of Solanum. In genera such as Triticum, plastids are present in both the generative cell and the sperm cell, but are not transmitted to the zygote. All three of these cases are examples of maternal or uniparental inheritance of plastids. Biparental inheritance, typical of Pelargonium, occurs when generative cell and sperm cell plastids are transmitted to the zygote.

In Pelargonium and Oenothera, the plastids of both parents can be detected in the progeny. In Pelargonium the genetic constitution of the parental plants can affect chloroplast inheritance in the progeny. In studies of crosses between plants containing both green and albino plastids (variegated plants), three groups of progeny are observed. One group of plants contains only green plastids, another only white plastids (lethal), and a third group is variegated. The ratio of plants in these groups is affected by the nuclear genotype of the maternal plant (Birky, 1983). However in Oenothera the ratio of progeny is independent of the maternal genotype. Thus, although the maternal genotype of some plant species can influence the ratio of progeny having parental plastid types, biparental inheritance of chloroplasts may still occur.

The genes controlling plastid inheritance can be nuclear or extranuclear, or a combination. Genes for many chloroplast polypeptides are encoded in both chloroplast and nuclear genomes and include ribulose-1,5-bisphosphate carboxylase, ATPase subunits, and components of the photosynthetic apparatus.

Chlamydomonas displays biparental chloroplast inheritance that is controlled by Mendelian alleles designated mt^+ (maternal) and mt^- (paternal). Each parental cell contains one large chloroplast. When the cells fuse, the mt^- chloroplast is eliminated, most likely through restriction endonuclease activity; the maternal (mt^+) chloroplast DNA is protected by methylation (Burton et al, 1979; Sager and Grabowy, 1983). Mutants have been obtained having partial methylation in both maternal and paternal cells (Bolen et al, 1982), however further methylation still occurs during gametogenesis in mt^+ but not mt^- cells (Sager and Grabowy, 1983). Such a methylation-restriction system is unlikely to be involved in chloroplast inheritance in higher plants, as no methylation has been detected in chloroplast DNA of higher plants.

One nuclear gene that controls chloroplast inheritance is iojap in maize. Wild type or heterozygous plants have fully green leaves and apparently normal plastids. Homozygous recessive plants are either lacking green pigment or are striped white on a green background. If a heterozygote is selfed, three-fourths of the progeny are phenotypically normal. The remaining fourth exhibits the iojap phenotype. This is characteristic of a single recessive nuclear gene. If these homozygous iojap plants

are crossed to a homozygous wild type female, the progeny are normal. If they are crossed to a homozygous wild type male, the progeny exhibit the iojap phenotype. Thus the trait is inherited maternally even though the nuclear genome encodes the wild type phenotype. The lack of plastid ribosomes is believed to cause this effect, not a mutation in the plastid DNA. Ribosomes are absent in plastids of plants exhibiting the iojap phenotype, but are present when the wild type nuclear genome is present. This indicates that a nuclear-encoded ribosomal protein is involved (Thompson et al, 1983).

Both nuclear and organellar genomes can thus be involved in organelle inheritance. Interactions between these genomes complicates the understanding of the events occurring during plant development. Plastids multiply by division (Butterfass, 1980). Plastid and nuclear DNA synthesis are not coupled, yet the number of chloroplasts per cell has been correlated with ploidy (Heinhorst et al, 1985; Butterfass, 1980). This implies that some coordination must be occurring. Both nuclear and plastid DNA synthesis depend on proteins synthesized in the cytoplasm, not in the plastid (Heinhorst et al, 1985). Thus plastid and nuclear replication are indirectly coordinated. This coordination can be tested by making different organellar combinations through somatic cell fusion.

Somatic Cell Fusion

As discussed above, most plants exhibit uniparental inheritance of organelles, typically due to exclusion of paternal organelles from the zygote. Somatic cell fusion allows the introduction of multiple genomes (nuclear, plastid, and mitochondrial) within a single cell. This produces novel combinations of genetic information that are excluded by uniparental inheritance or by barriers to sexual fertilization such as morphological or physiological incompatibilities. Somatic cell fusion can be used as a tool to study organelle inheritance in the limited number of plant species in which plant regeneration from protoplasts can be accomplished. Interactions between different organelles can then be studied during the process of regeneration. This process is straightforward in concept; isolated protoplasts are fused together, forming heterokaryons or heteroplasmons, according to whether the nuclear genomes are similar or dissimilar. These fusion products are then regenerated to full plants, which are subsequently examined to determine the relative contributions of the parental organelles to the progeny plant populations.

The genus Nicotiana has been used extensively for somatic cell fusion studies. This genus exhibits exclusively maternal inheritance in sexual hybridization in over sixty species. Most species of Nicotiana respond well in tissue culture and can be regenerated easily and in high frequencies from protoplasts. Thus